### What cichlids tell us about the social regulation of brain and behavior

Hans A. Hofmann\* and Russell D. Fernald, Neuroscience Program, Stanford University, Stanford, CA 94305, USA

#### **ABSTRACT**

Fishes are the largest vertebrate group and the object of considerable scientific attention. Cichlid fish, in particular, have attracted many researchers because of the species diversity and the complexity of their social behaviors. For decades, cichlid fish have played a prominent role in behavioral research, contributing to our understanding of the functions and causes of natural behavior. The mouthbrooding cichlid Haplochromis (Astatotilapia) burtoni (Günther) from the East African Lake Tanganyika lends itself to the study of social influences on the brain. Its complex, though easily observable behaviors and the occurrence of two distinct classes of males, those with territories and those without, facilitated insights into how physiological mechanisms interact with social behavior. For example, maturation of juveniles is suppressed in the presence of adult fish. Furthermore, the hypothalamic-pituitary-gonadal axis is shaped by social experience, as can be seen in the change of the size of gonadotropin-releasing hormone neurons after a change in social status. These neural and endocrine modifications are reversible, as each individual can change frequently between dominant and subordinate states depending on habitat stability. In addition, body growth depends on social status and immediate social history. This phenomenon can be explained in the context of life-history theory as a differential allocation of resources towards growth or reproduction, depending on social status. Putative causal factors of socially regulated growth include the neuropeptide somatostatin and growth hormone. The understanding of the social control of physiological processes gained in H. burtoni will be useful to understand the evolution and neurobiology of other social systems.

The social systems and sensory capacities of fishes have attracted diverse scientific attention for many reasons. Most often cited are two facts: First, there are an enormous number of fish species (>26,000) and second, fish occupy virtually every living space in the hydrosphere from alkaline springs to the Antarctic Ocean. A consequence of their extraordinary success is that fish species have evolved a remarkably diverse collection of behavioral adaptations. Among the teleosts, cichlids have attracted particular attention because their explosive speciation has produced a striking diversity of behavioral systems and corresponding sensory adaptations.

### Fishes as model systems for ethological research

A triumph of early ethologists was to transcend descriptive analysis by providing a framework for understanding the order underlying animal behavior. By studying particularly important life events such as feeding and reproduction in species with behavioral interactions less complex than ours, early ethologists Konrad Lorenz (1981) and Niko Tinbergen (1951) identified the central tenets of classical ethology,

namely the fixed action pattern and the key or sign stimulus. Fixed action patterns are complex behavioral acts, elicited by specific key stimuli and per-

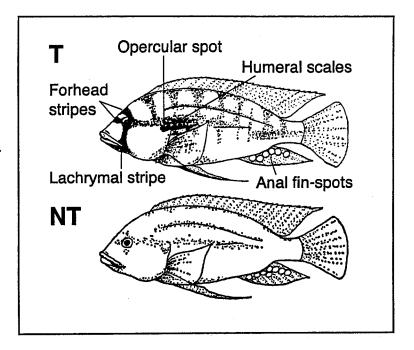
JOURNAL OF AQUARICULTURE AND AQUATIC SCIENCES

CICHLID RESEARCH: STATE OF THE ART

VOLUME IX

Page 1

Figure 1. Schematic illustration of the body patterns for typical territorial (T) and nonterritorial (NT) males. Top: the T male possesses distinctive anal fin spots and dark forehead and lachrymal stripes and is brightly colored, including orange humeral scales. The overall body color may be either yellow or blue. Bottom: NT males lack the robust markings of their territorial counterparts and are sand-colored overall. (Adapted from Fernald, 1977.)



formed by all or nearly all members of a species of a given age and sex. Though questions have been raised about how fixed such action patterns actually are (cf. Camhi, 1984), there is broad agreement that this categorization scheme remains useful for analyzing and interpreting animal behavior. Since many of the fixed action patterns were interpreted as stereotypical, ritualized displays that serve as communication signals among members of the same species, ethologists analyzed which features served to signal other animals. These motor patterns are often comprised of complex body movements as well as exposure of brightly colored patches on the body. Identification of the essential elements of the stimulus led to the idea of a sign or key stimulus (Tinbergen, 1951). A key stimulus is defined as a set of special features within a display that is part of the general stimulus pattern and responsible for eliciting particular behavioral actions.

Sign stimuli have been identified and studied in many species under a variety of conditions. Ethologists learned that many such stimuli and their behavioral responses could be studied effectively in the laboratory by constructing models on which elements of the stimulus could be presented. The most famous

of these models were constructed by Tinbergen (1948) to analyze the features causing territorial fights in the three-spined stickleback. Through systematic variation of parameters of the signal, Tinbergen (1948) showed that the color red, only when located on the belly of a dummy presented to a territorial male, caused neighboring animals to initiate a fight. This classic example showed that both the stimulus pattern (on the belly) and its context (territorial neighbor) determine whether the signal will produce a response. Sign stimuli can be presented in any modality, though the majority of signals that have been studied in detail are visual.

Discoveries of fixed action patterns and sign stimuli have provided significant insights about the proximal factors responsible for animals behaving as they do. However, these elemental features of behavioral activity need to be understood in the natural context of the animal. In real life, animals normally behave and interact continuously so that there is a seamless interrelationship among the fixed action patterns and sign stimuli. In 1954, Baerends and Baerends van Roon published a landmark monograph describing the behavior of numerous cichlid species. These remarkable animals clearly displayed attributes well suited for the then new field of ethology. First, within limits, they could be studied in the laboratory without compromising their natural behavior. Second, they were behaviorally active, making it possible to

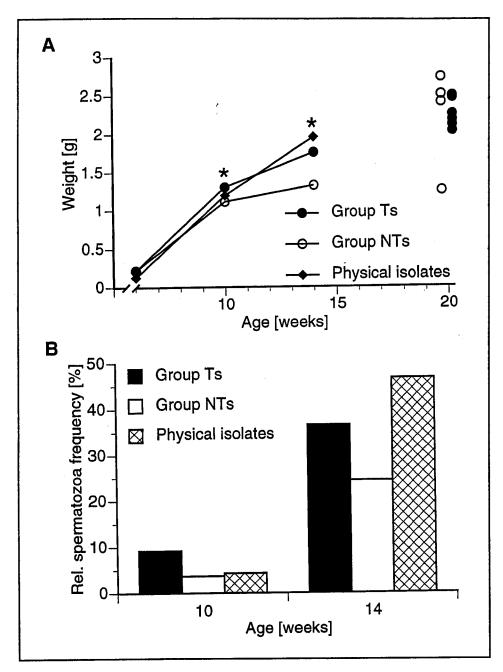
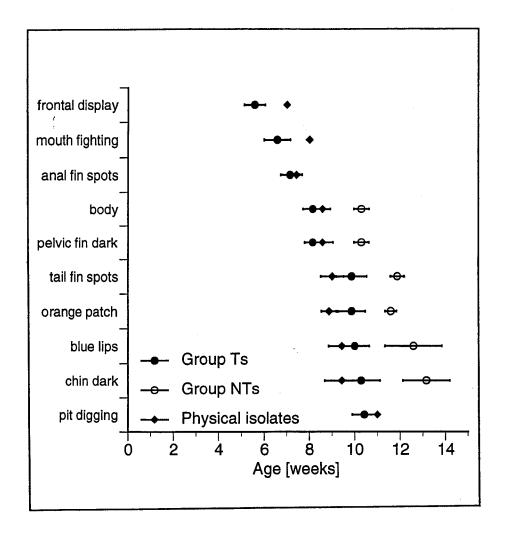


Figure 2. Development and maturation in group-reared (open and filled circles) and physically isolated (diamonds) juvenile *H. burtoni*. (A) Growth rates expressed as body weight for the different categories. Asterisks indicate that group-reared territorial fish (Ts, filled circles) weigh significantly more after 10 and 14 weeks as compared to their nonterritorial (NTs, open circles) tankmates. Differences in standard lengths are not significant (data not shown). Note that after 20 weeks size differences are no longer evident. (B) Relative estimates of mature spermatozoa in cross-sections of the central testicular lobule. Note the rapid increase in physically isolated males between week 10 and week 14. (After Fraley & Fernald, 1982; Davis & Fernald, 1990)

Figure 3. Age of first appearance of representative behavior and color patterns in H. burtoni males. Symbols for each social condition depict mean ± standard deviation of days when each pattern was first observed. Note that 0 is the release of fry from the mother's mouth following about two weeks of incubation. (Modified from Fraley & Fernald, 1982.)



collect quantifiable behavioral data in a reasonable amount of time. Third, there were sufficient numbers of species so that comparisons among closely related species were possible. Thus, cichlids offered a unique opportunity to examine both proximate mechanisms and ultimate functions of animal behavior in the same model system.

# HAPLOCHROMIS BURTONI: A MODEL SYSTEM FOR STUDYING SOCIAL INFLUENCES ON THE BRAIN

Over the last two decades, our laboratory has been using an African cichlid fish as a model system to discover how the visual system continues to function during rapid growth (see Fernald, 1984, for a review),

JOURNAL OF AQUARICULTURE AND AQUATIC SCIENCES
CICHLID RESEARCH: STATE OF THE ART
VOLUME IX
Page 4

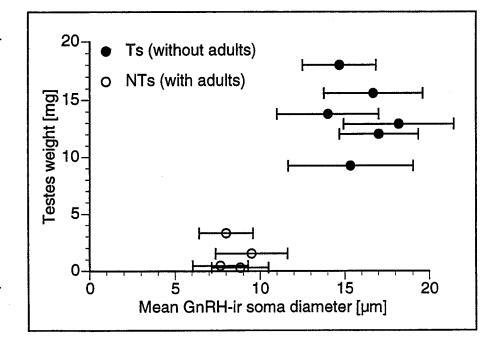
and to learn how social interactions lead to changes in the brain (see Fernald and White, 2000, for a review). Both of these lines of research depend on specific, unique characteristics of cichlids for the specific experiments, yet the insights obtained have proven generally applicable.

In the African cichlid fish, Haplochromis (Astatotilapia) burtoni (Günther), males exist in two different states which they can change reversibly: those with territories and those without (Fernald, 1977). Territorial (T) males are brightly colored, having a blue or yellow body color, a dramatic black stripe through the eye, vertical black bars on the body, a black spot on the tip of the gill cover and a large red patch just behind it. In contrast, nonterritorial (NT) males are cryptically colored and difficult to distinguish from the background and from females that are similarly camouflaged (Figure 1). Social communication among these fish depends primarily on visual signals (Fernald, 1984). In their natural habitat, the

shallow shorepools and river estuaries of Lake Tanganyika (Coulter, 1991), H. burtoni live in a leklike social system in which T males vigorously defend contiguous territories (Fernald and Hirata, 1977 a, b). The animals are very active, performing at least 19 distinct fixed action patterns during fast paced social encounters (Fernald, 1977). They spend considerable time digging a pit in the center of their territory, occasionally exchanging threat displays with their neighbors at their common territorial boundaries, frequently chasing NT animals away and soliciting and courting females. Solicitation and courtship behaviors are easily identified since the male displays bright coloration patterns towards the courted female. Courtship includes "leading" the female toward the territory and "courting", when the male quivers his opened, brightly colored anal fin in front of the female. Females led into the territory may feed by nipping at, and sifting through the bottom cover. NT males often mimic this behavior accurately enough so the T males will allow

them to enter the territories before the deception is discovered and the female impersonator is chased off. If the female responds to these entreaties by the male, he will lead her into his pit and continue the elaborate courtship movements, swimming to the front of the female and rapidly quivering his entire body with his spread anal fin in her view. The spawning male repeatedly interrupts his courtship and mating to chase off intruders. If sufficiently stimulated, the female lays her eggs at the bottom of the pit, collecting them in her mouth almost immediately. After she has laid several eggs, the male swims in front of her, again displaying the egglike spots on his anal fin (ocelli), his body quivering. The male displays this fin because the spots may seem to the female like eggs not yet collected (Wickler, 1962). So, while attempting to "collect" the spots, the female ingests the milt ejected near them by the male and ensures fertilization. After several bouts of this alternating behavior, the female may go to the territory of another male to lay more eggs or depart

Figure 4. Testes weights of 20 weeks old early-maturing (without adults present) territorial males (Ts; filled circles) and maturation-suppressed (with adults present) nonterritorial males (NTs; empty circles) plotted against the respective average soma diameters for the largest 30% of preoptic GnRH-IR neurons (± standard deviation). Neuron sizes are independent of body size in this experiment. Note the striking differences in cell size as well as testes weight between the two groups. (After Davis & Fernald, 1990.)



from the territorial arena with the fertilized eggs to brood them (Fernald, 1984).

This brief description of the natural behavior of H. burtoni reveals the extensive role of visual signals in social interactions and how much the social scene governs the behavior of individual animals. Each behavioral act influences the next, both in the observed individual and in the animals involved in the interaction. During the behavior, a great deal of information is being exchanged between individuals. What does the animal attend to and what are its intentions? Several specific fixed action patterns and key stimuli have been studied in this species. In particular, the role of the black eyebar has been studied in detail. Leong (1969) tested the response of T males to dummies with various configurations of the distinctive body patterns. Leong showed that the black eyebar (lachrymal stripe) alone increased the attack readiness toward blinded target fish, and the orangered patch of humeral scales alone decreased attack readiness. Subsequent experiments using similar dummy presentations revealed that the response was dependent on the alignment of the eyebar relative to the body and thus - since the eyebar angle in real fish is fixed - on the angular orientation of the animal in space (Heiligenberg and Kramer, 1972; Heiligenberg et al. 1972). Taken together, these experiments supported the idea that the black eyebar and the red humeral patch constitute key stimuli that cause behavioral responses of opposite types. When males were reared from hatching in complete isolation, they showed the same response to the presentation of dummies with an appropriately placed black eyebar and humeral patch (Fernald, 1980). The selective response to the appropriate key stimuli in naive animals suggests that the pattern recognition capability for these key stimuli is innate. Although a great deal about H. burtoni aggressive behavior is known already, a detailed analysis of sensory assessment cues, including but not limited to eyebar and humeral patch, based on game theory still remains to be done (cf. Enquist & Leimar, 1983; Enquist & Jakobsson, 1986).

In *H. burtoni*, the visual system has also been examined for evidence that these signals are impor-

JOURNAL OF AQUARICULTURE AND AQUATIC SCIENCES
CICHLID RESEARCH: STATE OF THE ART
VOLUME IX

Page 6

tant in the natural habitat (Fernald and Hirata, 1977a). In the primary habitat, shorepools and river estuaries along Lake Tanganyika, the color patterns on the body match the filtering properties of the water and appear to be optimized for the detectability of the relevant visual signals (Fernald and Hirata, 1977a). Moreover, the visual system itself is well matched to both the environment and the social signals. H. burtoni has a retina with three types of cone photoreceptors sensitive to distinct wavelengths (Fernald and Liebman, 1980), a prerequisite for trichromatic vision. Further analysis revealed that the photoreceptors are arranged in an array which is optimal for color vision (Fernald, 1981). The spectral sensitivity measured behaviorally (Allen and Fernald, 1985) showed that the animal could distinguish three wavelengths as predicted from the morphological measurements. Thus the visual system is matched to the environment and to the social signals produced by conspecifics including the red humeral patch and black eyebar.

## GROWTH AND THE SOCIAL CONTROL OF PHYSIOLOGICAL AND BEHAVIORAL PROCESSES IN *H. BURTONI*

Fish growth is indeterminate and can be rapid, especially in juveniles. To maintain a functioning visual system, new cells are added to the lens and retina continuously (Johns & Fernald, 1981; Fernald and Wright, 1983; Fernald, 1983, 1989). However, not all animals grow equally and most of the differences in growth rate depend on social conditions. Social control of maturation and growth is widespread among animals and takes a variety of forms depending on the species (Borowsky, 1973; Francis, 1988; Berglund, 1991; Schultz et al., 1991). In *H. burtoni*, we have begun to discover when social behavior regulates physiological processes, and some hints of the mechanisms responsible.

As young *H. burtoni* grow, the presence of conspecifics influences their behavioral and gonadal development and, even with food supplied *ad libitum*, their growth (Fraley and Fernald, 1982). For the first seven to eight weeks, group-living juveniles show the same growth as do broodmates reared in physical isolation (with visual contact; Figure 2). Although their standard lengths do not differ after this point in time, group-reared males that do not acquire and defend territories gain less weight than those with territories do, a difference that is not evident after 20 weeks

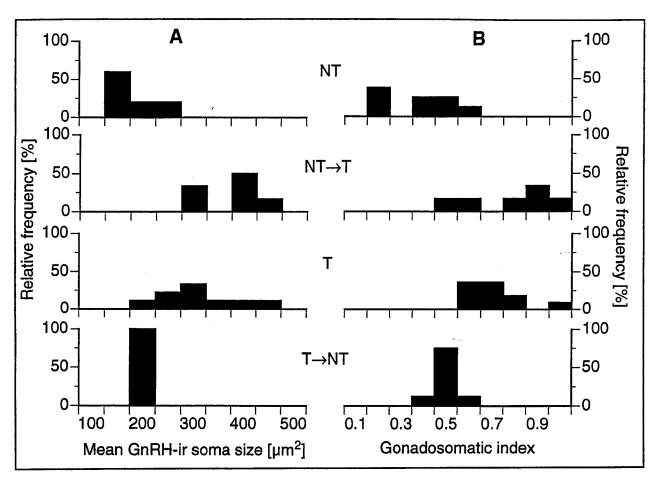
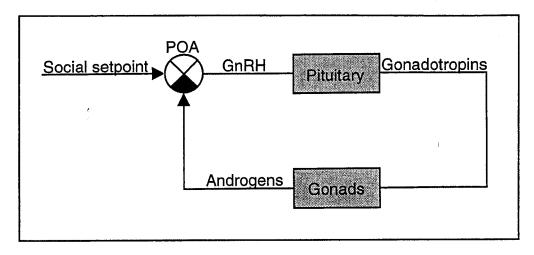


Figure 5. Frequency histograms of (A) mean soma sizes of preoptic area GnRH-IR neurons and (B) gonadosomatic indices (GSI). Percentage of individuals are plotted for each social category. There are significant differences, in soma sizes as well as GSI, between Ts and ascended NT→Ts when compared to NTs and descended T→NTs. (Modified after Francis et al., 1993.)

(Figure 2A). In addition, group-reared males that establish territories develop their gonads faster than group-reared NT fish and animals reared under isolation (Figure 2B). After 14 weeks, however, when territories are well established, physically isolated males, who all display aggressive behaviors, tend to possess more spermatozoa and be larger than Ts in the group condition, possibly a result of less energy expended by isolates due to the lack of physical competition (Figure 2). Concomitantly, group-reared fish show early developing agonistic/aggressive behavioral patterns (chase, tailbeat, fin spread) and chromatic patterns (eyebar, opercular spot) more than two weeks before their corresponding appearance in animals reared in physical isolation (Figure 3). However, later developing behaviors (frontal display, mouth fighting, pit digging) are first displayed by isolates at

almost the same time as by group-raised Ts (Figure 3).

Interestingly, some of the distinctive color patterns, including anal fin spots, body patterns, pelvic fin dark, tail fin spots, orange patch, blue lips, and dark chin can also be observed simultaneously or even earlier in isolates (Figure 3). Since NTs are not prevented from feeding by Ts (Fraley & Fernald, 1982), it seems that certain behaviors associated with territoriality (e.g., chasing and biting) are sufficient to suppress their maturation. In a social system where territorial space is limited (in the wild, only about 30% of the adult males are territorial and thus reproduc-



**Figure 6.** Feedback control model of GnRH regulation in male *H. burtoni*. Neurons in the preoptic area integrate both social and hormonal signals to regulate GnRH release. In this model, the setpoint for the GnRH level is determined by social signals and the maintenance of the GnRH level at this setpoint is achieved by negative feedback from gonadal androgens. (Modified from Soma et al., 1996.)

tively active) the regulation of maturation appears to be an adaptive solution to a limited resource.

The social regulation of growth and maturation does not only occur early in development. Juvenile males raised with adults present - the condition they usually encounter in the field - show suppressed gonadal maturation relative to those reared without adults (Davis and Fernald, 1990). As well as having smaller testes, these animals have smaller gonadotropin-releasing hormone (GnRH)-containing neurons in the preoptic area, a region in the ventral telencephalon adjacent to the hypothalamus (Figure 4). These neurons project to the pituitary (Bushnik & Fernald, 1995) where they release GnRH. The somata differ eightfold in volume depending on the social conditions. Since GnRH is the main signaling peptide that regulates reproductive maturity, the social control of maturation acts by changing structures in the brain.

Social status determines the physiology of the reproductive state, even in adult fish. This was demonstrated by changing males from T to NT or vice versa (Francis et al., 1993). To do this, T males were moved into communities with larger T males, where they became NT (T $\rightarrow$ NT). Correspondingly, NT males were moved to new communities consisting of

females and smaller males which they could dominate, and they became T (NT $\rightarrow$ T). In each case, after the subjects had remained in the altered social setting for four weeks, the size of GnRH containing cells was measured. Mean of both soma size of the preoptic area GnRH-immunoreactive (GnRH-IR) neurons (Figure 5A) and gonadosomatic index (GSI) (Figure 5B) were significantly larger in both NT→T and control T males than in T→NT and NT males. In two other GnRH-IR cell groups, one located in the terminal nerve region, the other in the mesencephalon, there was no difference in mean soma sizes between T and NT males (Davis & Fernald, 1990). This indicates that the status-linked variation in soma size is not a general property of GnRH containing neurons, but is, rather, confined to the preoptic area population (Davis & Fernald, 1990). The same result has been shown for GnRH mRNAs using in situ hybridization (White et al., 1995). Interestingly, female H. burtoni do not exhibit a similar kind of response to social interactions but do have modifications in their brain structures dependent on their reproductive status alone (White and Fernald, 1993).

As a consequence of the social control of GnRH, the social scene affects many other endocrine factors such as gonadotropins and sex steroids. For example, the amount of androgen released from the gonads depends on the social status of the individual (Soma et al., 1996). In *H. burtoni*, castrated T males' GnRH neurons do hypertrophy (Francis et al., 1992a; Soma

JOURNAL OF AQUARICULTURE AND AQUATIC SCIENCES CICHLID RESEARCH: STATE OF THE ART VOLUME IX
Page 8

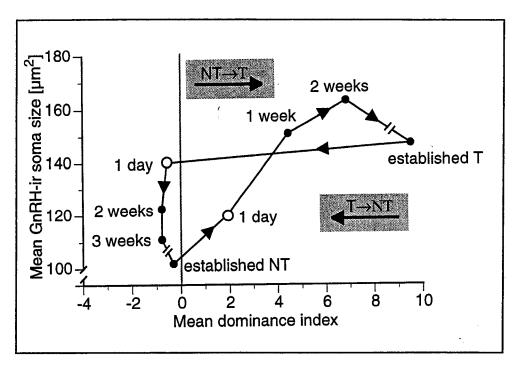


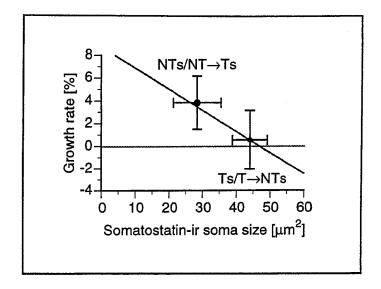
Figure 7. Mean two-dimensional GnRH-IR neuronal soma size plotted against the frequency of aggressive and submissive behaviors expressed as a Dominance Index (sum of aggressive acts minus sum of escape events / 3 min observation interval). Note the hysteresis like function as social status changes are asymmetric in regard to behavior and soma size: Although the behavioral change in  $T\rightarrow NT$  males is significantly faster (ca. 1 day) than in  $NT\rightarrow T$  males (ca. 2 weeks), the latter achieve soma sizes equivalent to those of Ts in one week, while  $T\rightarrow NT$ s require three weeks for their neurons to shrink to NT sizes. Empty circles indicate cases where soma size is hypothetical. Variances not shown for clarity. (Data from Nguyen et al., in preparation.)

et al., 1996). This observation led to the hypothesis that testosterone may negatively feed back onto the GnRH neurons, thereby limiting their maximum size as determined by territorial status. Social status appears to regulate the effect of androgen feedback, suggesting that there is a social setpoint for this feedback regulation (Figure 6). Interestingly, although the frequency of aggressive acts is reduced in castrated T males, they are able to maintain their social dominance (Francis et al., 1992b).

When examining the temporal dynamics of GnRH-neuron size changes after a change in social status, White et al. (in preparation) found that behavior as well as cell size changes are asymmetrical (Figure 7). Although fish who just lost territorial status (T \rightarrow NT) stop displaying aggressive behaviors immediately, the GnRH-containing neurons in the preoptic area do not shrink to NT size until three weeks after defeat. Conversely, ascending males (NT \rightarrow T) increase their aggressive displays relatively slowly over two weeks,

while the GnRH neurons reach T size only one week after a victory. The significance of this hysteresis in neural and behavioral change may be explained as the consequence of a life in an unstable world where reproductive opportunities may arise quickly for NTs (see below). After a defeat, switching to subordinate behaviors instantaneously helps the loser to prevent injuries. However, a new chance to establish a territory may soon arise, making the maintenance of an active reproductive physiology for a few weeks an adaptive strategy. Concomitantly, a newly ascended T should mature sexually as quickly as possible (even if his territorial behavior remains cautious for more than a week), since he may lose his territory sooner rather than later. Social status clearly determines both

Figure 8. Growth rates plotted as a function of the mean somatostatin-IR soma size in *H. burtoni*. NTs and NT $\rightarrow$ T males (filled circle; mean  $\pm$  standard deviations) have smaller soma cross-sectional areas and grow faster than Ts and T $\rightarrow$ NTs (filled diamond; mean  $\pm$  standard deviations). A linear regression analysis results in y = 0.19 \* x + 8.95, with  $r^2 = 0.4163$  (p < 0.001; n = 18). (After Hofmann & Fernald, 2000.)



soma size of preoptic area GnRH-IR neurons and GSI, and these effects are reversible. The relatively large testes and GnRH-IR neurons characteristic of T males are a consequence of their social dominance, and when this dominance advantage is lost, both neurons and testes shrink (Francis et al., 1993). It remains unknown exactly how social information is transformed into changes in the brain. There is, however, some evidence that visual information may be used to signal the state of individual animals.

Both the production and detection of visual signals have been subject to natural as well as sexual selection in cichlid fishes (e.g., Seehausen et al., 1997). In H. burtoni, the black lachrymal stripe across the eyes that signals aggression can appear and disappear within seconds, independently of other coloration patterns. This eyebar is controlled by a specialized branch of the VIth cranial nerve, which regulates localized changes in the pigment concentration (Muske and Fernald, 1987a). Incidentally, there is a striking difference in the sensitivity of the neural system controlling the eyebar between nonterritorial and territorial animals (Muske and Fernald, 1987b). Eyebars located on T males show much greater sensitivity to norepinephrine, the neurotransmitter that controls changes in expression of that social signal. Moreover, there is a long-term morphological change in the eyebar of T males. In such males, the eyebar tissue adds iridiphores that enhance the contrast and hence effectiveness of the signal (Muske and Fernald, 1987b). Thus, the production of this visual display is distinctly different at the physiological level in T males when compared to NT males. Understanding how visual information exchanged in a social context is transduced into structural changes in the brain is a major challenge. It seems likely that these processes will be common to many species, particularly to those where social interactions regulate reproduction. New insights will require combining observations of social interactions with physiological measurements.

While the mechanisms through which social signals regulate reproductive physiology are beginning to be understood, researchers are also learning more about the effects of the environment on social systems (see Lott, 1982, for a general review). We have recently shown in H. burtoni that habitat complexity influences the fraction of the male population that can sustain territories (Hofmann et al., 1999a). Moreover, the stability of the habitat affects duration of territorial tenure (Hofmann et al., 1999a). In a fluctuating habitat, where the three-dimensional layout changes frequently, males hold territories for a significantly shorter time period than in a stable habitat. The same is true for the duration of nonterritorial status. In an unstable world, such as the shorepools of Lake Tanganyika, the regulation of reproduction, growth and development appears adaptive. Only a fraction of the males can breed at any time and these animals appear to be particularly vulnerable to avian predators (Fernald and Hirata, 1977b), and hence territorial ownership may be relatively brief. In addition, winds and the presence of large animals can cause considerable environmental change (Fernald & Hirata, 1977b). As a consequence, reproductive opportunities may arise as frequently as they vanish. This phenomenon helps to explain the asymmetry found in the responses of GnRH neurons to changes in social status (see Figure 7).

Surprisingly, even a stable habitat results in a significant level of social change (Hofmann et al., 1999a). This intrinsic instability is caused by differential growth rates. Specifically, NTs and NT→Ts grow faster than Ts and T→NTs (Figure 8). This result may seem unexpected when compared to the weight differences between juvenile Ts and NTs, as discussed above. However, it may well be that during the weeks after territory establishment, juvenile Ts are comparable to adult NT→Ts, allocating their energy simultaneously towards reproduction and growth in order to gain a competitive advantage over other Ts. Conversely, juvenile NTs around 10 to 14 weeks of age may be similar to adult T→NTs, showing suppression of both growth and gonadal maturation as a consequence of social defeat. When does the switch in growth patterns occur? As noted above, at the age of 14 weeks group-reared Ts lose their developmental advantage when compared to broodmates reared in physical isolation, and many behaviors as well as coloration patterns appear simultaneously in both groups. According to the available data (Davis & Fernald, 1990), the size differences between Ts and NTs are no longer evident at the age of 20 weeks (Figure 2A). It may well be that these developmental shifts reflect the increased amount of energy established Ts have to spend for territory maintenance and reproductive effort. As a consequence, the social pressure on NTs may decrease, giving them opportunity to grow faster, which ultimately helps them to gain territoriality.

A possible mechanism regulating differential growth is the control of somatostatin release in the pituitary, where this neurohormone inhibits the release of growth hormone (GH; Brazeau et al., 1973; Gillies, 1997). As we have shown recently (Hofmann & Fernald, 2000), somatostatin-containing neurons in the preoptic area change size when social status and, consequently, growth rate change (Figure 8). The somata of these neurons are significantly larger in Ts and T-NTs as compared to NTs and NT-Ts. It is

unknown whether larger neurons produce more somatostatin to be released into the pituitary, or whether they represent an accumulation of somatostatin as its release is inhibited. Preliminary evidence from measurements of circulating GH (Hofmann et al., 1999b) hints that the latter may be the case, thus inhibiting the release of GH from the pituitary in NTs and NT $\rightarrow$ Ts. This surprising result brings into play a possible social regulation of insulin-like growth factor 1 (IGF-I) which mediates many of the somatic effects of GH and whose release is controlled by GH (Mommsen, 1998). Clearly, more data are needed here to unravel the complex interactions of neural and endocrine factors.

Why do animals that have lost a territory  $(T \rightarrow NTs)$ slow down their growth rate and even shrink? Behavioral stressors may play a role. As shown by Fox et al. (1997) in H. burtoni, status switches in both directions can be accompanied by elevated levels of the major stress hormone cortisol with the T-NT change showing the most pronounced increase. NT-T fish with increased cortisol levels usually did not maintain territoriality. Fish descending in rank consistently showed high levels of cortisol. Cortisol may be elevated by losing a territory and may in turn cause somatic growth to be downregulated. As has been shown in another cichlid, the tilapia Oreochromis mossambicus, chronic administration of cortisol leads to a reduction in body weight and reproductive parameters like gamete size and levels of sex steroids (Foo & Lam, 1993). Although the regulatory interactions between GH and cortisol are very complex (Thakore & Dinan, 1994; van Weerd & Komen, 1998, for critical reviews), in vivo experiments have demonstrated an inhibitory effect of glucocorticoids on somatic growth in many vertebrates including fish (e.g., Pickering, 1990).

Could cortisol also be involved in the growth rate differences between established Ts and NTs? Fox et al. (1997) showed that cortisol levels in Ts and NTs do not differ as long as the fish community remains unstable. However, in a situation of relatively high social stability, Ts have significantly lower levels of circulating cortisol than NTs. Under such a stable

situation NTs still grow faster than Ts. Therefore, growth may not be effectively inhibited by cortisol in those animals. Rather, we hypothesize that other factors may become significant when animals maintain a particular social behavior for many weeks (e.g. feeding habits, behavioral activity, energy expenditure).

### H. BURTONI: LESSONS FOR UNDERSTANDING OTHER SOCIAL SYSTEMS?

In their natural habitat, *H. burtoni* have limited territorial space, so there may be a selective advantage for males that are prepared for reproductive opportunities, adjusting their behavior and physiology quickly to changes in their situation.

The modulation of the brain by behavior makes sense in an evolutionary framework where the behavioral phenotype is seen as the locus of selective pressure. Phenotypic plasticity allows *H. burtoni* to reversibly adapt its behavior and physiology to changing social opportunities, thus allocating resources between reproduction and growth (Williams, 1966). Understanding how the evolution of life histories shapes the nervous system provides a unique chance to discover the universal mechanisms of social interactions. From the comparative aspect of evolutionary neurobiology, the astonishing diversity of cichlid fishes in the African Great Lakes and South America may help us to unravel the intricate interrelationships between habitat structure, behavior, and the brain.

#### **ACKNOWLEDGMENTS**

We thank A. Ettinger, A. Greenwood, and W. Loher for comments on earlier versions of the manuscript. The research was supported by a postdoctoral fellowship from the Deutsche Forschungsgemeinschaft (HAH) and by NIH-NS 34950 (RDF).

of

#### REFERENCES

Allen, E. E., and R. D. Fernald. 1985. Spectral sensitivity of the African cichlid fish, *Haplochromis burtoni*. Journal of Comparative Physiology A 157:247-253.

Baerends, G. P., and J. M. Baerends van Roon. 1950. An introduction to the study of the ethology of cichlid fishes. Behavior, Supplementum 1:1-235.

Berglund, A. 1991. Egg competition in a sex-role reversed pipefish: subdominant females trade reproduction for growth. Evolution 45:770-774.

Borowsky, R. 1973. Social control of adult size in males of *Xiphophorus variatus*. Nature 245:332-335.

Brazeau, P., W. Vale, R. Burgus, N. Ling, M. Butcher, J. Rivier, and R. Guillemin. 1973. Hypothalamic polypeptide that inhibits the secretion of immunoreactive pituitary growth hormone. Science 179:77-79.

Bushnik, T. L., and R. D. Fernald. 1995. The population of GnRH-containing neurons showing socially mediated size changes project to the pituitary in a teleost, *Haplochromis burtoni*. Brain, Behavior & Evolution 46:371-377.

Camhi, J. M. 1984. Neuroethology. Nerve Cells and the Natural Behavior of Animals. Sinauer, Sunderland, MA.

Coulter, G. W. 1991. Lake Tanganyika and Its Life. Oxford University Press, Oxford. Davis, M. R., and R. D. Fernald. 1990. Social con-

trol of neuronal soma size. Journal of Neurobiology 21:1180-1188.

Enquist, M., and S. Jakobsson. 1986. Decision making and assessment in the fighting behaviour of *Nannacara anomala* (Cichlidae, Pisces). Ethology 72:143-153.

Enquist, M., and O. Leimar. 1983. Evolution of fighting behaviour: decision rules and assessment of relative strength. Journal of theoretical Biology 102:387-410.

- Fernald, R. D. 1977. Quantitative observations of *Haplochromis burtoni* under semi-natural conditions. Animal Behavior 25:643-653.
- Fernald, R. D. 1980. Response of male cichlid fish, *Haplochromis burtoni*, reared in isolation to models of conspecifics. Zeitschrift für Tierpsychologie 54:85-93.
- Fernald, R. D. 1981. Chromic organization of a cichlid fish retina. Vision Research 21:1749-1753.
- Fernald, R. D. 1983. Neural basis of visual pattern recognition in fish. p. 569-580 in Advances in Vertebrate Neuroethology. (Ewert, J. P., R. R. Capranica, and D. J. Ingle, eds). Plenum Press, New York.
- Fernald, R. D. 1984. Vision and behavior in and African cichlid fish. American Scientist 72:58-65.
- Fernald, R. D. 1989. Retinal rod neurogenesis. p. 31-42 in Development of the Vertebrate Retina. (Finley, B. L., and D. R. Sengelaub, eds). Plenum Press, New York
- Fernald, R. D., and N. R. Hirata. 1977a. Field study of *Haplochromis burtoni*: quantitative behavioural observations. Animal Behavior 25:964-975.
- Fernald, R. D., and N. R. Hirata. 1977b. Field study of *Haplochromis burtoni*: habitats and co-habitants. Environmental Biology of Fishes 2:299-308.
- Fernald, R. D., and P. A. Liebman. 1980. Visual receptor pigments in the African cichlid fish, *Haplochromis burtoni*. Vision Research 20:857-864.
- Fernald, R. D., and S. A. White. 2000. Social control of brains: from behavior to genes. p. 1193-1208 in The Cognitive Neurosciences, 2<sup>nd</sup> Edition. (Gazzaniga, M. S., ed). MIT Press, Cambridge.
- Fernald, R. D., and S. E. Wright. 1983. Maintenance of optical quality during crystalline lens growth. Nature 301:618-620.

- Foo J. T. W., and T. J. Lam. 1993. Retardation of ovarian growth and depression of serum steroid levels in the tilapia *Oreochromis mossambicus* by cortisol implantation. Aquaculture 115:133-143.
- Fox, H. E., S. A. White, M. H. F. Kao, and R. D. Fernald. 1997. Stress and dominance in a social fish. Journal of Neuroscience 17:6463-6469.
- Fraley, N. B., and R. D. Fernald. 1982. Social control of developmental rate in the African cichlid, *Haplochromis burtoni*. Zeitschrift für Tierpsychologie 60:66-82.
- Francis R C. 1988. Socially mediated variation in growth rate of the midas cichlid: the primacy of early size differences. Animal Behaviour 36:1844-1845.
- Francis, R. C., B. Jacobsen, J. C. Wingfield, and R. D. Fernald. 1992a. Hypertrophy of gonadotropin releasing hormone-containing neurons after castration in the teleost, *Haplochromis burtoni*. Journal of Neurobiology 23:1084-1093.
- Francis, R. C., B. Jacobsen, J. C. Wingfield, and R. D. Fernald. 1992b. Castration lowers aggression but not social dominance in male *Haplochromis burtoni* (Cichlidae). Ethology 90:247-255.
- Francis, R. C., K. Soma, R. D. Fernald. 1993. Social regulation of the brain-pituitary-gonadal axis. Proceedings of the National Academy of Sciences USA 90:7794-7798.
- Gillies, G. 1997. Somatostatin: the neuroendocrine story. Trends in Pharmacological Sciences 18:87-95.
- Heiligenberg, W., and U. Kramer. 1972. Aggressiveness as a function of external stimulation. Journal of Comparative Physiology and Psychology 77:332-340.

Heiligenberg, W., U. Kramer, and V. Schulz. 1972. The angular orientation of the black eye-bar in *Haplochromis burtoni* (Cichlidae, Pisces) and its relevance to aggressivity. Zeitschrift für vergleichende Physiologie 76:168-176.

Hofmann, H. A., M. E. Benson, R. D. Fernald. 1999a. Social status regulates growth rate: consequences for life-history strategies. Proceedings of the National Academy of Sciences USA 95:14171-14176.

Hofmann, H. A., P.-Y. Le Bail, and R. D. Fernald. 1999b. Social control of growth and growth hormone levels in an African cichlid fish. Society for Neuroscience Abstracts 25:866.

Hofmann, H. A., and R. D. Fernald. 2000. Social status controls somatostatin-neuron size and growth. Journal of Neuroscience, 20:1248-1252.

Johns, P. R., and R. D. Fernald. 1981. Genesis of rods in teleost fish retina. Nature 293:141-142.

Leong, C.-Y. 1969. The quantitative effect of releasers on the attack readiness of the fish *Haplochromis burtoni*. Zeitschrift für vergleichende Physiologie 65:29-50.

Lorenz, K. 1981. The Foundations of Ethology (Vergleichende Verhaltensforschung). Springer-Verlag, New York.

Lott, D. F. 1982. Intraspecific Variation in the Social Systems of Wild Vertebrates. Cambridge Studies in Behavioural Biology. Cambridge University Press, Cambridge.

Mommsen, T. P. . 1998. Growth and Metabolism. p.65-97 in The Physisology of Fishes (Evans, D. H., ed). CRC Press, Boca Raton, FL.

Muske, L. E., and R. D. Fernald .1987a. Control of a teleost social signal. I. Neural basis for differential expression of a color pattern. Journal of Comparative Physiology A 160:89-97.

Muske, L. E., and R. D. Fernald .1987b. Control of a teleost social signal. II. Anatomical and physiological specializations of chromatophores. Journal of Comparative Physiology A 160:99-107.

Pickering, A. D. 1990. Stress and the suppression of somatic growth in teleost fish. p. 473-479 in Progress in Clinical and Biological Research, Vol. 342: Progress in Comparative Endocrinology (Epple, A., C. G. Scanes, and M. H. Stetson) 473-479. Wiley-Liss, New York.

Schultz E. T., L. M. Clifton, and R. R. Warner. 1991. Energetic constraints and size-based tactics: The adaptive significance of breeding-schedule variation in a marine fish, *Micrometrus minimus* (Embiotocidae). American Naturalist 138:1408-1430.

Seehausen O., J. J. M. Van Alphen, and F. Witte. 1997. Cichlid fish diversity threatened by eutrophication that curbs sexual selection. Science 277:1808-1811.

Soma, K. K., R. C. Francis, J. C. Wingfield, and R. D. Fernald. 1996. Androgen regulation of hypothalamic neurons containing gonadotropin-releasing hormone in a cichlid fish: integration with social cues. Hormones & Behavior 30:216-226.

Thakore, J. H., T. G. Dinan. 1994. Growth hormone secretion: the role of glucocorticoids. Life Sciences 55:1083-1099.

Tinbergen, N. 1948. Social releasers and the experimental method required for their study. Wilson Bull 60:6-52.

Tinbergen, N. 1951. The Study of Instinct. Clarendon Press, Oxford.

van Weerd, J. H., and J. Komen. 1998. The effects of chronic stress on growth in fish: A critical appraisal. Comparative Biochemistry and Physiology 120:107-112.

White, S. A., and R. D. Fernald. 1993. Gonadotropin-releasing hormone-containing neurons change size with reproductive state in female *Haplochromis burtoni*. Journal of Neuroscience 13:434-441.

White, S. A., T. L. Kasten, C. T. Bond, J. P. Adelman, and R. D. Fernald. 1995. Three gonadotropin-releasing hormone genes in one organism suggest novel roles for an ancient peptide. Proceedings of the National Academy of Sciences USA 92:8363-8367.

Wickler, W. 1962. Zur Stammesgeschichte funktionell korrelierter Organ- und Verhaltensmerkmale: Eiattrappen und Maulbrüten bei afrikanischen Cichliden. Zeitschrift für Tierpsychologie 19:129-164.

Williams, G. C. 1966. Adaptation and Natural Selection. Princeton Univ. Press, Princeton, NJ.

|  |  |   | , |
|--|--|---|---|
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  | • |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |
|  |  |   |   |