Hair-compatible sponge electrodes integrated on VR headset for electroencephalography

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Abstract

Virtual reality (VR) technology has emerged as a promising tool for brain-computer interaction and neuroscience research due to its ability to provide immersive and interactive experiences for its users. As a powerful tool to noninvasively monitor the cortex, electroencephalography (EEG) combined with VR represents an exciting opportunity for the measurement of brain activity during these experiences, providing insight into cognitive and neural processes. However, traditional gel-based EEG sensors are not compatible with VR headsets, and most emerging VR-EEG headsets utilizing rigid comb electrodes are uncomfortable after prolonged wear. To address this limitation, we created soft, porous, and hair-compatible sponge electrodes based on conductive poly(3,4-ethylenedioxythiophene) polystyrene sulfonate/melamine (PMA) and integrated them onto a VR headset through a customized, flexible circuit for multichannel EEG during VR task performing. Our PMA sponge electrodes can deform to make contact with the scalp skin through hairs under the pressure naturally applied by the strap of the VR headset. The specific contact impedance was consistently below 80 kΩ·cm², even at hairy sites. We demonstrated the capability of our VR-EEG headset by recording alpha rhythms during eye closure at both hairless
and hairy sites. In another demonstration, we developed a VR task to evoke the contingent negative variation potential and achieved a classification accuracy of $0.66 \pm 0.07$, represented by the cross-validated area under the receiver operating characteristic curve. Our sponge-electrode-integrated VR headset is user-friendly and easy to set up, marking a step toward future reliable, comfortable, and reusable VR-EEG technology.

**Keywords:** PEDOT:PSS, soft electrode, electroencephalography, virtual reality, brain-computer interface

### INTRODUCTION

Electroencephalography (EEG) is a non-invasive technique that records neural activity from the scalp, offering high temporal resolution, affordability, and versatility compared to alternative modalities such as functional magnetic resonance imaging (fMRI) and invasive electrocorticography (ECoG). EEG has been widely used for various applications, including sleep monitoring, clinical diagnosis and treatment of neurological disorders such as epilepsy and stroke, and as a primary signal modality for both clinical and non-clinical brain-computer interfaces (BCI). In recent years, virtual reality (VR) technology has emerged as a new tool in the fields of cognition assessment, rehabilitation, pain relief, and BCI. VR can create controlled environments that integrate intuitive, immersive, and interactive elements with innovative input methods such as gaze direction and hand gestures, potentially replacing conventional visual stimulation and feedback techniques. Therefore, the integration of EEG and VR represents a promising opportunity for the improvement of both EEG and VR systems. EEG can provide a real-time stream of brain activity and cognitive state information that can be utilized by the VR application, while VR can provide a unique environment for evoking and studying brain activity in realistic and immersive simulations.

Current EEG and VR systems are implemented separately in hardware, resulting in cumbersome and complicated systems when they are simply combined. Emerging research has demonstrated the viability of integrating EEG electrodes directly on VR headsets for simultaneous EEG recording and VR stimulation, but these studies primarily target hairless areas such as the forehead. However, regions of interest for brain activity analysis and many BCI applications are often located underneath the hairy parts of the scalp such as the motor cortex and visual cortex, which are hard to access by existing VR-EEG systems. To overcome hair interference in conventional EEG, the conductive liquid gel is commonly used, but it is time-consuming to set up, limited in operating time, uncomfortable, and requires trained personnel. Additionally, some ingredients in the gel electrodes, such as propylparaben, have been found to be harmful to the skin. To address these issues, paintable gel electrodes with a fast liquid-to-solid transition speed have been developed, simplifying the scalp preparation and electrode application process and reducing the required time compared to commercial gel electrodes. However, these electrodes require removal by washing off after use, limiting their integration with other systems for multiple uses.

Dry electrodes have gained attention for EEG due to their ease of use, setup, removal, and integration with wearable systems compared to gel electrodes. Various dry electrode designs, including microneedle, nanotube, and pillar electrodes, have been explored over the past decade. These electrodes can penetrate hair bundles to make direct contact with the scalp, providing reasonable EEG recordings. However, their fabrication processes are often complex microfabrication processes, which are high-cost and time-consuming. Furthermore, transdermal microneedles can cause pain and potential skin infections. Recent studies have found that soft electrodes made of elastomer pillars coated with conductive materials, such as gold and poly(3,4-ethylenedioxythiophene) polystyrene sulfonate (PEDOT:PSS), have greatly reduced stiffness and improved skin compatibility. However, an additional
adhesive layer is necessary to prevent the delamination of the coating layer under pillar deformation, which adds complexity and cost to the fabrication process.

Soft conductive sponges offer a hair-friendly option for EEG recording, as they can make better contact with the scalp when compressed despite hairs\cite{29}. These sponge electrodes can be integrated onto various types of headwear such as caps\cite{30} and headbands\cite{31}, making them ideal for portable and wireless EEG recording. Recent works reported that silver-nanowire-coated melamine (MA) sponges can record high-fidelity steady-state visual evoked potentials (SSVEP) on hairy scalps\cite{29,32}. However, silver nanowires require a polymer coating for improved chemical stability and reduced toxicity. PEDOT:PSS is a conducting polymer with high chemical stability, conductivity, and biocompatibility\cite{33,34}. It has both ionic and electronic conductivity, enabling lower contact impedance with the skin than that of gold\cite{35}. These characteristics make PEDOT:PSS a popular choice for electrode materials in bioelectrical signal recording\cite{29,33,34,36}. In this paper, we fabricate soft PEDOT:PSS/MA (PMA) sponge electrodes that can be easily integrated on a commercial VR headset for EEG recording during VR experiences. The PMA sponge electrode is mechanically compliant and can conform to the hairy scalp upon being pressed by the VR headset strap, resulting in high-fidelity multichannel EEG recordings. We developed a VR program to demonstrate the PMA sponge electrode-based VR-BCI system. We have successfully detected the contingent negative variation (CNV)-related potentials using our VR-EEG BCI system.

EXPERIMENTAL

Chemicals and materials
MA sponges were purchased from Amazon. PEDOT:PSS (Clevios PH1000, solid content 1.3%) was obtained from Heraeus Precious Metals North America Daychem LLC (Vandalia, OH, USA). Dimethyl sulfoxide (DMSO, 99.9%) was purchased from Beantown Chemical (Hudson, NH, USA). Copper films were purchased from Nimrod Copper Company. Solid gel electrodes (Kendall ECG electrodes) and comb electrodes were obtained from Kendall and OpenBCI (Brooklyn, NY, USA), respectively. All chemicals were used as received without further purification.

Fabrication of PMA sponges
PMA sponges were prepared by a dipping method. In brief, MA sponge pieces were punched into cylinders with a height of 10 mm and a diameter of 12 mm. A PEDOT:PSS solution was prepared by mixing 7 g of Clevios PH1000 dispersion and 3 g of DMSO under stirring at room temperature for 30 minutes. The MA sponge cylinders were then dipped into the PEDOT:PSS solution and squeezed four times to enhance infiltration of the PEDOT:PSS solution. The PEDOT:PSS solution with MA sponges was then placed into a desiccator and kept under vacuum for 15 minutes to further enhance infiltration of the PEDOT:PSS solution. The sponges were then scooped up, and excess PEDOT:PSS solution was removed using filter paper. The sponges were dried at 110 °C in an oven for 2 hours. After another dipping and drying process, PMA composite sponges with a dark blue color were obtained.

Characterization
The morphology of both MA sponges and PMA sponges was characterized by scanning electron microscopy (SEM, Scios 2HiVac, Thermal Fisher Scientific, Canada). The UV-Vis absorption of PEDOT:PSS solution and washing solution was measured on a UV-Vis NIR spectrometer (Cary 5000, Agilent, USA). The mechanical properties of the sponges were characterized using a dynamic mechanical analyzer (RSA-G2, TA Instruments).
Impedance measurement
Before impedance measurement, the forehead and mastoid areas (A1 and A2) were cleaned with Nuprep skin preparation gel. PMA sponge electrodes were placed on the specified locations according to the standard 10-10 EEG system, and two solid gel electrodes were placed on A1 and A2 as the reference and ground electrodes. The contact impedance spectrum was measured on the Autolab PGSTAT204 (Metrohm, Switzerland) in the range of 1 Hz to 1000 Hz and the Brain Vision Recorder software (Brain Products GmbH, Munich, Germany) at 15 Hz.

EEG recording and analysis
EEG signals were recorded from PMA sponge electrodes integrated on the Meta Quest 2 headset, fitted with a Meta Quest 2 Elite Strap (Meta, California, USA). All analysis was performed using custom MATLAB scripts and functions from EEGLAB[37].

Eye close vs. open comparison
EEG signals were recorded using the Brain Vision LiveAmp 32 amplifier and the Brain Vision Recorder software (Brain Products GmbH, Munich, Germany) and analyzed using custom MATLAB scripts. The experimenter recorded triggers on the Brain Vision Recorder software while giving instructions to the subject. The EEG signals were initially filtered using a bandpass filter ranging from 1.5 Hz to 50 Hz, and the alpha band was defined as 8 to 12 Hz.

VR experiment analysis
Three healthy male subjects aged 23-28 participated in the CNV recording experiment. The VR task was designed and implemented in Unity (editor version 2020.3.11f1) and was conducted using the Meta Quest 2 headset and controllers. The grand average analysis and decoding analysis were performed using signals from the Cz channel in MATLAB. The signal was preprocessed by bandpass filtering 0.1 to 1 Hz using a fourth-order Butterworth filter. Baseline correction was applied using a 100 ms window prior to Stimulus 1. Trials with an absolute sample value exceeding 50 μV were defined as artifactual and rejected from further analysis. For the decoding analysis, the EEG time samples between the two stimuli (-4.3 s before Stimulus 2 to Stimulus 2 onset) were extracted after bandpass filtering from 0.01 to 3 Hz. The extracted samples were used as features for the decoder after min-max normalization to values between 0 and 1. The decoding accuracy was computed using leave-one-out block-wise cross-validation from five blocks, using diagonal linear discriminant analysis (LDA).

RESULTS AND DISCUSSION
The design of our sponge electrode integrated VR headset for simultaneous EEG recording and VR display is illustrated in Figure 1A. The sponge electrodes were integrated onto the VR headset using a custom flexible connector array (FCA) that runs along the headset straps. Once the user wears the VR headset, the electrodes, and FCA are concealed from view. The FCA provides a soft and stable electrical connection between the electrodes and the recording system, and it is also compatible with the VR headset. In contrast, previous VR headset-integrated EEG electrodes typically rely on rigid stud connectors that have to puncture the strap to establish a connection with the recording system[17]. Moreover, the FCA enables the sponge electrodes to be distributed across various scalp locations, allowing EEG recording from multiple brain regions, including frontal, frontocentral, central, centroparietal, and occipital regions. Notably, the headset straps can apply pressure on the sponge electrodes, enabling them to deform and make better contact with the hairy scalp, thereby reducing electrode impedance. Figure 1B shows a photograph of the PMA sponges, which are 12 mm in diameter and 10 mm in height. Figure 1C shows a photograph of the FCA, comprising nine copper O-ring connectors with seamless copper traces. The entire system is fabricated on a 25 μm-thick polyethylene terephthalate (PET) substrate that can bend to conform to the headset straps, ensuring
Figure 1. Design of our VR-EEG headset. (A) A schematic illustration of the PMA sponge electrodes integrated on a commercial VR headset to make good contact with the hairy scalp for EEG. The blue dash lines labeled on the VR headset strap indicate the outline of the FCA laminated on the other side of the strap; (B) A photograph of the PMA sponge electrodes; (C) A photograph of the FCA that connects all sponge electrodes to the back of the VR headset for connection to the data acquisition system; (D) An exploded view illustrating the installation of one PMA sponge on the VR headset through insertion into the open holes of the FCA, which is adhered to the strap by a double-sided elastomeric sticker; (E) The VR headset with nine PMA sponge electrodes integrated on it. EEG: electroencephalography; FCA: flexible connector array; PMA: poly (3,4-ethylenedioxythiophene) polystyrene sulfate/melamine; VR: virtual reality.

stable integration. The topside of the connectors and traces are insulated with Kapton tape to prevent direct copper-skin contact. The connector has an inner diameter of 10 mm, slightly smaller than that of the sponge, allowing it to deform and fit snugly into the connector hole for a secure connection. The FCA was fabricated using a laser cutting process [Supplementary Figure 1], and the details of the fabrication process are provided in the supporting information. A 1 mm-thick elastomer double-sided sticker was utilized to further secure the sponge electrodes to the VR headset straps, providing stability under shear force [Figure 1D]. Figure 1E displays the VR headset equipped with nine integrated sponge electrodes, which is ready to be worn by the user for simultaneous EEG recording and VR display.
The PMA sponge was prepared using a dipping process. Figure 2A shows a schematic illustration of the fabrication process. The MA sponge cylinders were dipped in a PEDOT:PSS solution diluted with DMSO in a vacuum system to facilitate the infiltration of PEDOT:PSS solution into the pores of sponges. After removing the excess solution, the sponges were dried in an oven to evaporate the remaining solvents. The dip coating was repeated to increase the PEDOT:PSS coating and conductivity. After coating, the MA sponge changed from white to dark blue [Figure 2B], indicating successful PEDOT:PSS loading. The dried PEDOT:PSS layer is stable and non-dissolvable in water[38]. After washing the PMA sponge in saline solution ten times, no visible PEDOT absorption was detected in the UV-vis spectrum within the range of 500-800 nm[39]. However, a distinct PEDOT absorption was observed in a PEDOT:PSS solution with a low concentration of 0.02 wt.% [Supplementary Figure 2]. This result shows that the PMA sponge is inert, stable, and capable of functioning under sweaty conditions without any additional protection layer. SEM images in Figure 2C-E compare the structure of the MA sponge before and after PEDOT:PSS coating. The original MA sponge has a porous structure with a MA fiber skeleton. After the dipping process, in addition to the coating on the MA fiber skeleton, there are PEDOT:PSS flakes suspending on adjacent MA skeletons due to the evaporation of the remaining solution within the sponge [Figure 2D]. The width of the PEDOT:PSS flakes ranges in hundreds of micrometers, and the thickness of the flakes is around hundreds of nanometers. It was observed that some MA fibers with the skeletal structure exhibited fractures subsequent to the coating of PEDOT:PSS [Figure 2E]. This could potentially be attributed to the squeezing process during the fabrication of the PMA sponge. The PMA sponge is compressible [Figure 2F]. Therefore, we studied the mechanical properties of the PMA sponge and the MA sponge under different compressive strains from 30% to 90%. As shown in Figure 2G and H, the PMA sponge and the MA sponge have similar stress-strain curves at 30% and 50% strains, except for the lower Young’s modulus of the PMA sponge (approximately 54.6 kPa) compared to the MA sponge (approximately 102.8 kPa) in the 0%-5% regime. We hypothesize that the partially fractured skeleton within the PMA sponge contributes to its reduced stiffness. The maximum compressive stresses of the PMA sponge at compressive strains of 70% and 90% are 33.58 kPa and 265.38 kPa, respectively, which are higher than those of the MA sponge (27.11 kPa and 83.21 kPa, respectively) due to the PEDOT:PSS flakes formed within the pores in the sponge [Figure 2D and E]. We further investigated the durability of the PMA sponge by performing consecutive loading-unloading up to a compressive strain of 50% for 20 cycles. As shown in [Supplementary Figure 3], the PMA sponge reaches similar maximum compressive stresses in every cycle, but hysteresis in stress-strain exists. The conductivity of the PMA sponge can be adjusted by modifying the dipping process and PEDOT:PSS solution concentration. The resistance of the PMA sponge prepared with one dipping is 85.62 ± 25.42 Ω, while that of the sponge prepared with two dips is 5.6 ± 2.2 Ω [Supplementary Figure 4A]. The resistance of the PMA sponge decreased from 33.2 ± 6.3 Ω to 5.6 ± 2.2 Ω as the PEDOT:PSS concentration increased from 0.39% to 0.91% [Supplementary Figure 4B]. Furthermore, the resistance of the PMA sponge can be further reduced by applying pressure, which increases the conductive path due to sponge compression [Supplementary Figure 5]. This is advantageous for the sponge electrode as it enhances the conductivity under the pressure of the VR headset strap during EEG recording.

Contact impedance plays a vital role in brain activity recordings for EEG electrodes. Therefore, we integrated the PMA sponges onto a VR headset and measured the contact impedance on both hairless and hairy sites. Using a calibrated, highly sensitive, and flexible hybrid-response pressure sensor (HRPS) developed in our group [Supplementary Figure 6][40], we have quantified that the VR headset strap can apply a pressure of 9.48-11.70 kPa on the hairless site and 10.62-12.21 kPa on the hairy site. The pressure applied by the strap can induce deformation of 25.41%-33.90% on dry sponges and 44.32%-47.71% on wet sponges. This deformation leads to a further decrease in the resistance of the sponges. The equivalent circuit model of the electrode on the hairless site is illustrated in Figure 3A. The sponge electrode is modeled as a parallel resistance (Re) and capacitance (Ce). As the electrode directly contacts the skin, the electrode-skin interface...
is modeled as a contact resistance (Rc) and capacitance (Cc), which is determined by the gap between the electrode and the skin. The epidermis and dermis are modeled as a resistor-capacitor (RC) circuit ($R_{Ep}$, $C_{Ep}$) and a resistor ($R_d$), respectively. The softness of the sponge electrode allows it to form good contact with the skin and minimize the gap, leading to a small Rc and large Cc. We measured the contact impedance of the sponge electrodes at the Fp1 and Fp2 locations while wearing the VR headset across three subjects. The results are plotted in Figure 3B, with the error bars indicating the standard deviation. The full frequency spectrum collected by the electrochemical workstation (Autolab PGSTAT204) shows that the sponge electrode has an average specific impedance of $12.73 \pm 4.67$ kΩ-cm$^2$ at 10 Hz, while that for a solid gel electrode (Kendall ECG electrodes) is $3.79 \pm 2.53$ kΩ-cm$^2$. The variation in impedance is attributed to the diverse skin conditions among the subjects. Given the chemical stability of our sponge electrodes, we further investigated their impedance under wet conditions on the same subject. The wet sponge was prepared by adding several drops of deionized water to the sponge and squeezing out the excess liquid. The specific contact impedance of the wet sponge obtained by the Brain Vision Recorder software is $6.41 \pm 0.58$ kΩ-cm$^2$ at 15 Hz, which is three times lower than that of the dry electrode ($19.40 \pm 3.60$ kΩ-cm$^2$ at 15 Hz) [Figure 3C]. One possible reason for the reduced impedance under wet conditions is that the wetting
**Figure 3.** Contact impedance of the sponge electrode on hairless skin (A-C) and hairy skin (D-F). (A) Cross-sectional schematic and equivalent circuit model of the sponge electrode on hairless skin; (B) Electrode-skin impedance spectrum of the sponge electrode and solid gel electrode on a total of nine hairless sites of three different subjects. The markers are average values, and the error bars indicate the standard deviation; (C) A comparison of the contact impedance between the dry and wet sponge electrodes on hairless skin (n = 6). The inset plots the loading-unloading stress-strain curves of dry (black) and wet (red) sponges; (D) Cross-sectional schematic and equivalent circuit model of the sponge electrode on hairy skin; (E) Electrode-skin impedance spectrum of the sponge electrode and solid gel electrode on the hairy scalp; (F) A comparison of the contact impedance between the dry and wet sponge electrodes on the hairy sites (n = 6); (G) Electrode-skin contact impedance spectrum of the sponge electrode before (black) and after (red) being compressed 100 times (three measurements at each frequency); (H) The contact impedance of the sponge electrodes on different locations is stable after 60 min of continuous wear; (I) A radar chart that compares the sponge electrodes with other EEG electrodes in the literature based on the impedance, softness, hair compatibility, and reusability criteria. Grading is based on the following criteria. Impedance lower than 10 kΩ·cm² is good, 10 kΩ·cm²–100 kΩ·cm² is medium, and higher than 100 kΩ·cm² is poor. Softness with Young’s modulus lower than 50 kPa is good, 50 kPa–1 MPa is medium, and higher than 1 MPa is poor. Hair compatibility, if the electrode cannot deform to contact the hairy scalp, it is considered poor; if the electrode can deform to reach the hairy scalp but has medium impedance, it is defined as medium; if the electrode can form very good contact with the hairy scalp with good impedance, it is considered high. Reusability: if the electrode can be reused without being destroyed after use, it is considered good; otherwise, it is considered poor. EEG: electroencephalography.

The process makes the sponge softer, leading to a change of Young’s modulus from approximately 54.6 kPa to 30.0 kPa [inset, Figure 3C]. This change in modulus allows the sponge to deform more easily when pressed by the headset strap, resulting in better electrode-skin contact and an increase in Cc. Another possible reason is that the stratum corneum has an improved conductivity when wet[41], contributing to a lower impedance with a reduced REp.

We further studied the contact impedance of sponge electrodes on the hairy scalp by integrating two sponges on the O1 and O2 locations of the VR headset. The equivalent circuit model of the sponge on the
hairy site is similar to that on the hairless site, except that the hair creates an additional barrier between the electrode and the skin. The electrode-hairy skin interface is modeled as an RC circuit of \(R_{ch}\) and \(C_{ch}\), where \(R_{ch}\) and \(C_{ch}\) are the contact resistance and the contact capacitance, respectively [Figure 3D]. The full frequency spectrum shows that the dry sponge electrodes have a high specific impedance of 452.71 k\(\Omega\)·cm\(^2\) at 10 Hz due to the separating effect of the hair. By contrast, when wet, the specific impedance of the sponge decreased to 45.50 k\(\Omega\)·cm\(^2\) at 10 Hz, which is approximately ten times lower than that of the dry sponge [Figure 3E]. Impedance measurement by the Brain Vision Recorder software showed that the dry sponge on the hairy site has a specific impedance of 464.71 ± 72.24 k\(\Omega\)·cm\(^2\) at 15 Hz, while the wet sponge has a much lower specific impedance of 65.54 ± 12.67 k\(\Omega\)·cm\(^2\) at 15 Hz [Figure 3F]. These results further highlight the importance of the softness of sponges in establishing a good interface with the hairy scalp and reducing impedance. In addition, since no skin preparation was done on the hairy site, the stratum corneum on the hairy scalp is thicker, resulting in a more significant increase in conductivity compared to the forehead.

The mechanical stability of the sponge under repeated compression is essential for reusability. As shown in Figure 3G, the specific impedance of the sponge remains unchanged after 100 cycles of squeezing at 50% compressive strain, demonstrating that the sponge electrodes can be used multiple times with the VR headset. The electrochemical stability of the electrode over a prolonged period is crucial for long-term EEG signal recording. Therefore, we measured the stability of electrode impedance in different locations for 60 minutes. The contact impedance varied depending on the location of the electrode, with the dry sponge on the forehead exhibiting lower impedance compared to the wet sponges on the hairy scalp [Figure 3H]. Most of the electrodes exhibited stable impedance over time. Furthermore, we investigated the reusability of the PMA sponge. As shown in Supplementary Figure 7, the electrode exhibited similar impedance after putting on and taking off the VR headset ten times, indicating that our sponge electrode can be used for at least ten uses. Finally, we compared our sponge electrodes with other types of electrodes in the literature, including wet gel, textile, comb electrodes, and soft pillar electrodes. We can see that our sponge electrode outperforms those electrodes with softness, low impedance, good compatibility with hair, ease of integrability, and reusability [Figure 3I, Table 1]. In comparison to other sponge electrodes [Table 1][17,29,42], our PMA sponges were simply but reliably integrated onto the VR headset through an ultrathin FCA, making the whole system easy to assemble and also soft and comfortable to wear.

Next, we studied the capability of the sponge electrodes to record EEG signals during eye-close and eye-open conditions, using four participants in our study. The purpose of this experiment is to demonstrate that sponge electrodes can successfully record increased alpha rhythm, which is a characteristic of EEG that is present during the absence of visual stimuli, such as the eyes being closed. The sponge electrodes were integrated onto the VR headset using a flexible FCA at Fp2 as a hairless site and Cz and Oz as hairy sites [Figure 4A]. The end of the FCA can be directly connected to the EEG data acquisition system without requiring additional fixation [Figure 4B]. As shown in Figure 4C, the filtered EEG signals from Cz and Oz demonstrated a clear increase in amplitude in the alpha band (8-12 Hz) during the eye-close period compared to the eye-open period, resulting in increased power. Time-frequency analysis and power spectra density (PSD) results confirmed this increase in alpha power in the range of 8-12 Hz during the eye-closed period compared to the eye-open period [Figure 4D and E]. Thus, the EEG data recorded with the sponge electrode exhibited the characteristic feature of increased alpha oscillation power during the absence of visual stimulation[43]. The stable integration of the sponge electrodes on the VR headset enables consistent EEG recording over a typical VR session of 60 minutes [Supplementary Figure 8], which is sufficient considering that typical VR applications do not exceed this duration to avoid issues such as VR cybersickness and discomfort[42]. We also compared the performance of our sponge electrodes with commercial electrodes on both hairless and hairy sites. For hairless sites, we attached a solid gel electrode at
## Table 1. Summary of the performance of different types of hairy-compatible EEG electrodes

<table>
<thead>
<tr>
<th>Materials</th>
<th>Fabrication</th>
<th>Softness</th>
<th>Hair compatibility</th>
<th>Specific Impedance (kΩ·cm²)</th>
<th>Connection</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>PMA sponge</td>
<td>Dip coating</td>
<td>Yes</td>
<td>Yes</td>
<td>19.40 ± 3.60 (15 Hz, forehead) 65.54 ± 12.67 (15 Hz, hairy site)</td>
<td>FCA</td>
<td>This work</td>
</tr>
<tr>
<td>Gelatin gel</td>
<td>Chemical polymerization</td>
<td>Yes</td>
<td>Yes</td>
<td>5.46 ± 0.76 (10 Hz, hairy site)</td>
<td>Stud</td>
<td>Wang et al. [22]</td>
</tr>
<tr>
<td>PEDOT hydrogel</td>
<td>Chemical polymerization</td>
<td>Yes</td>
<td>Yes</td>
<td>20.7 (10 Hz, hairy site)</td>
<td>Stud</td>
<td>Hsieh et al. [9]</td>
</tr>
<tr>
<td>PEDOT:PSS-coated PDMS pillars</td>
<td>Replica and coating</td>
<td>Yes</td>
<td>Yes</td>
<td>--</td>
<td>--</td>
<td>Zhang et al. [28]</td>
</tr>
<tr>
<td>Au-coated silicon pin</td>
<td>Microfabrication and thermal deposition</td>
<td>Yes</td>
<td>Yes</td>
<td>7.5 (10 Hz, hairy site)</td>
<td>--</td>
<td>Wang et al. [23]</td>
</tr>
<tr>
<td>Cellulose sponge</td>
<td>--</td>
<td>No</td>
<td>Yes</td>
<td>1171.3 (forehead); 1089-1727.9 (hairy site)</td>
<td>Stud</td>
<td>Ko et al. [42]</td>
</tr>
<tr>
<td>Conductive fabric and metal-coated sponge</td>
<td>--</td>
<td>Yes</td>
<td>No</td>
<td>18.1-28.3 (15 Hz, forehead); 53.1-91.5 (15 Hz, hairy site)</td>
<td>Stud</td>
<td>Kuang et al. [18]</td>
</tr>
<tr>
<td>Ag NW/MA sponge</td>
<td>Dip coating</td>
<td>Yes</td>
<td>Yes</td>
<td>0.6-1.2 (30 Hz, hairy site)</td>
<td>Silver wire</td>
<td>Lin et al. [29]</td>
</tr>
</tbody>
</table>

FCA: flexible connector array; MA: melamine; PMA: poly (3,4-ethylenedioxythiophene) polystyrene sulfate/melamine; PEDOT-PSS: poly(3,4-ethylenedioxythiophene) polystyrene sulfonate

the Fp1 location, which had similar skin conditions to the neighboring Fp2 [Supplementary Figure 9A]. Our sponge electrode demonstrated similar EEG features to the solid gel electrode, with a Pearson’s correlation coefficient (r) of 0.977 between the EEG signals recorded by the two electrodes [Figure 4F]. On hairy sites, our sponge electrode performed comparably to a commercial comb electrode located adjacent to it near Cz [Supplementary Figure 9B], with a Pearson’s correlation coefficient (r) of 0.860 [Figure 4G]. It is worth noting that the sponge electrode is compatible, soft, and skin-friendly, not causing any skin irritation and not leaving visible markings after wearing for 1 hour, whereas the comb electrode is stiff and uncomfortable, leaving noticeable marks on the skin [Supplementary Figure 10]. The ease of setup, high stability, and user-friendliness of our sponge-electrode-integrated VR headset makes it a promising system for simultaneous EEG recording and VR interaction.

Finally, we evaluated the performance of our sponge electrode in a VR-BCI system. To this end, we designed a CNV task in a custom first-person perspective VR game for the “Go/No-Go” classification. The electrode was placed on Cz, located over the central region of the cerebral cortex [Figure 5A], which is a typical location of interest for detecting CNV potentials. CNV is a well-established slow cortical EEG potential associated with anticipation and attention generated from sources in the prefrontal and central regions of the cerebral cortex [44-46]. Our VR game was designed to resemble decision-making while driving, which is a commonly used scenario and application for CNV-based BCIs [Figure 5B, Movie S1] [47-49]. The subject was instructed to remember the direction of the middle symbol of the Flanker task (Stimulus 1) [Figure 5C] [50] and respond exactly 4.3 seconds later (as guided by the countdown on display) by pressing a button on the VR controller the moment the diverging sign (Stimulus 2) appeared when the Stimulus 1 was a “Go”. If Stimulus 1 was a “No-Go”, the subject was instructed to ignore Stimulus 2. The input was only accepted within a 300 ms window after Stimulus 2, and the response time of the subject was displayed as feedback during the “Go” trials to keep the subject alert to Stimulus 2 [Figure 5B]. Positive visual feedback in the form of a gem object and an increasing score was delivered if the response of the subject was correct.
In the VR-BCI experiment, three subjects completed five blocks of the VR task while EEG was simultaneously recorded using the sponge electrode. Each block consisted of 6 “Go” and 6 “No-Go” trials. Grand average analysis of the EEG signals showed that a negative potential slope was developed between the two stimuli only for “Go” trials [Figure 5D], which is consistent with the result in previous reports[46]. Leave-one-out block-wise cross-validation was performed, resulting in a cross-validated area under the receiver operating characteristic curve (AUC) of $0.66 \pm 0.07$ for the three subjects [Figure 5E].
CONCLUSIONS

In this paper, we developed soft and conductive PMA sponge electrodes that can be integrated with a commercial VR headset for simultaneous EEG recording and immersive VR stimulation. The sponge electrodes can form sufficient contact with both hairless and hairy scalps for high-quality EEG recordings. In addition, the sponge electrode is compressible and can maintain a stable contact impedance with the skin after 100 cycles of compression, which enhances the durability and longevity of the electrode. Our sponge-electrode-integrated VR headset allowed for stable measurement of EEG features, such as alpha rhythms, for 60 minutes. Furthermore, we have demonstrated that our electrodes are capable of recording VR-evoked CNV potentials. Our VR-EEG system is easy to set up, reliable for EEG recordings during VR interaction, user-friendly, customizable, and has the potential for use in various applications requiring simultaneous EEG recording and VR interaction. In the future, through custom-designed VR headset straps and FCA, more sponge electrodes can be placed strategically for EEG recordings from desirable brain regions depending on the application, such as recording other event-related potentials, sensorimotor rhythms, etc., for the purposes of emotion recognition or workload evaluation. Moreover, VR headsets that can conform to the scalp and provide consistent pressure could significantly enhance the quality of recorded EEG during even longer periods of time.
DECLARATIONS

Authors’ contributions
Conceptualization, electrode preparation, methodology, data acquisition, analysis, and interpretation, writing original draft: Li H
Methodology, data acquisition, software and data analysis, manuscript revising: Shin H, Zhang M, Huh H, Riveira N, Kwon G
Supervision, funding acquisition, and manuscript revision: Sentis L
Project discussion, supervision, and manuscript revision: Millán JdR
Conceptualization, funding acquisition, project administration, resources, supervision and writing, review, and editing: Lu N

Availability of data and materials
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Conflicts of interest
All authors declared that there are no conflicts of interest.

Ethical approval and consent to participate
The study was conducted in accordance with the ethical guidelines and principles set forth by the Internal Review Board (IRB) of the University of Texas at Austin under approval number STUDY00004005. All participants were informed about the experimental procedure and signed the informed consent forms prior to participation.

Consent for publication
Not applicable.

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Supplementary Information

Hair-Compatible Sponge Electrodes Integrated on VR Headset for Electroencephalography

1. Fabrication of FCA and sponge electrode integration onto VR headset

The patterns of the FCA circuits were designed by AutoCAD and produced by a laser cutter (U4, LPFK Laser & Electronics). The process involved the following steps:

1. A 25 μm-thick PET film was covered with a double-sided polyimide (PI) tape with a PET liner.
2. The PET/PI/PET film was cut into FCA outline patterns with 10 mm-diameter holes at the recording sites, and the excess was peeled off to get the FCA outline.
3. The top PET liner was removed, and a 25 μm-thick copper film was attached to the exposed PI double-sided tape.
4. Another 25 μm-thick Kapton tape was added as an insulation layer on top of the copper film.
5. The film was cut into the FPC patterns, and the excess was peeled off to get the FCA with insulated 1 mm-wide copper traces.
2. Supplementary Figures

**Figure 1.** Schematic of the fabrication process of the FCA.

**Figure 2.** UV-vis spectrum of the washing solution and a highly diluted PEDOT:PSS solution.
Figure 3. Stress-strain curve of the PMA sponge recorded under repeated loading and unloading at 50% strain for 20 cycles.

Figure 4. Resistance of the PMA sponges prepared with different parameters. (A) Comparison of the PMA sponges with different dipping cycles ($n = 12$). (B) Comparison of the PMA sponges prepared in PEDOT:PSS solution with different PEDOT:PSS concentrations ($n = 12$).

Figure 5. The decrease of the sponge resistance with the increase of compressive strain.
Figure 6. The setup used in measuring the pressure exerted by the VR headset strap on the hairless site (A) and on the hairy site (B).

Figure 7. The comparison of the contact impedance of sponge electrodes on the forehead before and after ten uses.
Figure 8. (A) Alpha bandpass filtered EEG signals (8-12 Hz) captured during eye-close and eye-open periods after 60 min. (B) Time-frequency analysis of the EEG signals recorded at Oz. Colors show event-related spectral perturbation (ERSP), calculated as the ratio of average amplitudes in the experimental condition (eyes-close or eye-open) to the baseline epoch (taken from the start of the recording before subject instructions were given) after 60 min. In (A) and (B), the shaded areas indicate an eye-closed period. (C) PSD of the EEG signals recorded at Oz under eyes-close (blue) and eye-open (red) conditions after 60 min.

Figure 9. Photographs of the commercial solid gel electrode (A) and comb electrode (B) used for comparison during EEG recording.
Figure 10. The comparison of the skin wearing medical tape fixed sponge electrode (left) and comb electrode (right) for 1 h.